

Vitamin D in pregnancy and lactation: maternal, fetal, and neonatal outcomes from human and animal studies¹⁻⁴

Christopher S Kovacs

ABSTRACT

During pregnancy and lactation, mothers require significant amounts of calcium to pass on to the developing fetus and suckling neonate, respectively. Given the dependence of adult calcium concentrations and bone metabolism on vitamin D, one might anticipate that vitamin D sufficiency would be even more critical during pregnancy and lactation. However, maternal adaptations during pregnancy and lactation and fetal adaptations provide the necessary calcium relatively independently of vitamin D status. It is the vitamin D-deficient or insufficient neonate who is at risk of problems, including hypocalcemia and rickets. Due to poor penetrance of vitamin D and 25-hydroxyvitamin D [25(OH)D] into milk, exclusively breastfed infants are at higher risk of vitamin D deficiency than are formula-fed infants. Dosing recommendations for women during pregnancy and lactation might be best directed toward ensuring that the neonate is vitamin D-sufficient and that this sufficiency is maintained during infancy and beyond. A dose of vitamin D that provides 25(OH)D sufficiency in the mother during pregnancy should provide normal cord blood concentrations of 25(OH)D. Research has shown that during lactation, supplements administered directly to the infant can easily achieve vitamin D sufficiency; the mother needs much higher doses (100 μ g or 4000 IU per day) to achieve adult-normal 25(OH)D concentrations in her exclusively breastfed infant. In addition, the relation (if any) of vitamin D insufficiency in the fetus or neonate to long-term nonskeletal outcomes such as type 1 diabetes and other chronic diseases needs to be investigated. *Am J Clin Nutr* 2008;88(suppl):520S–8S.

INTRODUCTION

Calcium and bone metabolism in adults depend heavily on concentrations of vitamin D and its active metabolite 1,25-dihydroxyvitamin D [1,25(OH)₂D]. Without 1,25(OH)₂D, the body cannot absorb calcium and phosphorus adequately, secondary hyperparathyroidism supervenes, the skeleton loses mineral content (secondary osteoporosis), and new bone is not adequately mineralized (rickets or osteomalacia) (1). Hypocalcemia can occur, but secondary hyperparathyroidism supports blood calcium through skeletal resorption.

During pregnancy and lactation, mothers provide large amounts of calcium to the developing fetus and suckling neonate, respectively (2, 3). Given that adult calcium and bone metabolism depend on vitamin D sufficiency, vitamin D sufficiency would seem to be especially critical during pregnancy and lactation. However, as this review shows, maternal adaptations during pregnancy, lactation, and fetal development provide the necessary calcium relatively independently of vitamin D. It is only

after birth that dependency on vitamin D becomes evident, at least with respect to calcium metabolism and skeletal health.

Due to the relative paucity of data obtained during human pregnancy and lactation, this review includes discussions of animal data on vitamin D's role in mammalian calcium metabolism. Studies in humans should confirm all pertinent findings from animal models, but this might never be possible for certain aspects of pregnancy and fetal development.

To avoid an unduly lengthy reference list, I direct the reader to a 1997 comprehensive review by Kovacs and Kronenberg (2), with >550 primary references on the issues discussed here, and several recent reviews that cite studies published since 1997 (3–8).

ADAPTATIONS DURING PREGNANCY

During gestation, the human fetus accretes 30 g Ca on average, of which 99% is contained within the skeleton. More than 150 mg/kg of this calcium is actively transferred each day across the placenta during the third trimester.

Serum calcium concentrations (which include ionized, protein-bound, and complexed fractions) fall early in pregnancy as a result of the drop in serum albumin. This artifact of pregnancy's hemodilution is physiologically unimportant and is not evidence of true hypocalcemia. Ionized calcium concentrations, the physiologically important fraction, do not change during pregnancy. Parathyroid hormone (PTH), as measured by "intact" assays, falls to the lower end of the normal range and can become undetectable in North American and European women (no studies have used the newer "bio-intact" PTH assays). In contrast, studies of women from Gambia, Asia, and other areas where calcium and vitamin D intake are low have found that PTH concentrations do not drop during pregnancy. Levels of other hormones with potential calcium-regulating effects—including estradiol, prolactin, placental lactogen, and the calcium-regulating hormone parathyroid hormone-related protein (PTHrP)—increase during pregnancy.

¹ From the Faculty of Medicine–Endocrinology, Memorial University of Newfoundland, St John's, Newfoundland, Canada.

² Presented at the National Institutes of Health conference "Vitamin D and Health in the 21st Century: an Update," held in Bethesda, MD, September 5–6, 2007.

³ Supported by the Canadian Institutes of Health Research, the National Sciences and Engineering Research Council of Canada, and the Dairy Farmers of Canada.

⁴ Address reprint requests to C Kovacs, Health Sciences Centre, 300 Prince Philip Drive, St John's, Newfoundland A1B 3V6 Canada. E-mail ckovacs@mun.ca.

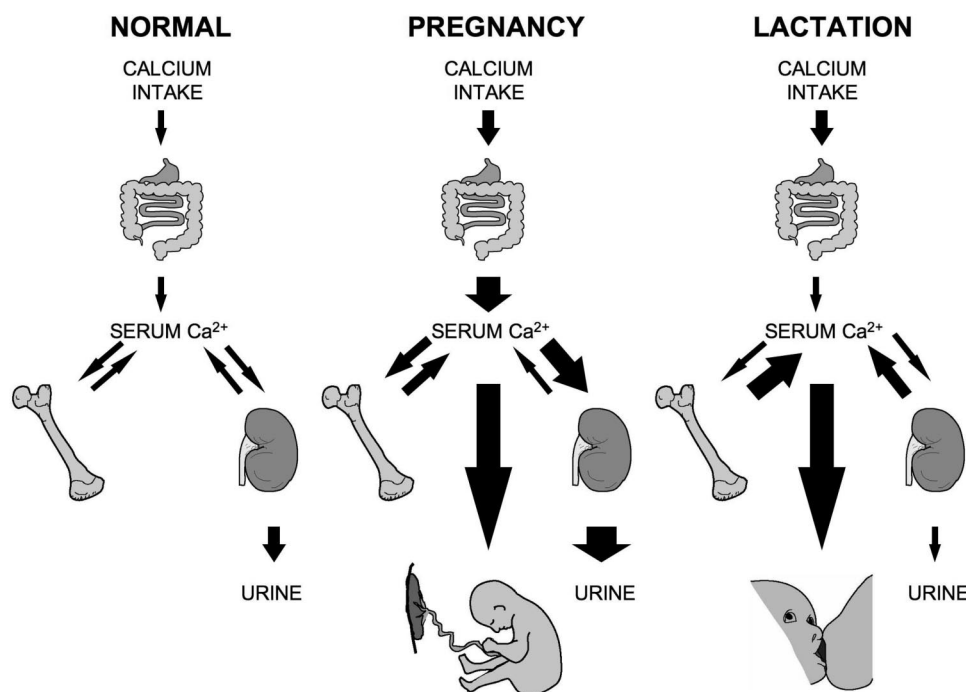


FIGURE 1. Schematic illustration contrasting calcium homeostasis in human pregnancy and lactation compared with normal. Arrow thickness indicates a relative increase or decrease with respect to the normal, nonpregnant state. Adapted from reference 2, copyright 1997, The Endocrine Society.

The pregnancy-induced adaptations to maternal calcium homeostasis are illustrated in **Figure 1**. Doubling the rate or efficiency of intestinal calcium absorption starting early in pregnancy appears to meet the fetal need for calcium. Skeletal resorption can also provide mineral to the circulation, but evidence is mixed on whether the maternal skeleton contributes substantial amounts of calcium to the fetus. Bone resorption markers are modestly increased during pregnancy (less than during lactation), and bone biopsies from women at the time of first-trimester abortions show histomorphometric evidence of increased bone resorption. Older, longitudinal studies of bone mineral density (BMD) during pregnancy (using single and dual-photon absorptiometry) showed no change in BMD. Studies using the more modern dual-energy X-ray absorptiometry 3–9 mo before planned pregnancies and 1–6 wk after delivery showed no change or a 1–4% decrease in BMD at the spine or hip between the first and second measurements. Serial ultrasound measurements at the heel have shown apparent BMD loss during pregnancy, but such peripheral measurements might have little relevance to the clinically and physiologically important content of the spine and hip. The maternal kidneys do not reclaim calcium avidly during pregnancy; instead, urinary calcium excretion increases in parallel with the increase in intestinal calcium absorption.

Rats and mice have a much shorter gestation period (22 and 19 d, respectively) and transfer 95% of their calcium to 8–12 fetuses per litter during the last 4–5 d of gestation. Ionized calcium levels are stable until late pregnancy, when they can drop during the rapid calcium transfer to the fetus. PTH is suppressed early in pregnancy but can increase in late pregnancy. Skeletal mineral content increases starting early in pregnancy, at least in certain strains of rats and mice.

VITAMIN D METABOLISM DURING PREGNANCY

25-Hydroxyvitamin D [25(OH)D], the storage form of vitamin D, readily traverses the hemochorial placentas of rats (9) and probably crosses the hemochorial human placenta readily, such that cord blood 25(OH)D concentrations are equal to or up to 20% lower than maternal concentrations (10, 11). Thus, for neonates to be born with adult-normal 25(OH)D concentrations, their mothers must be vitamin D–sufficient. Passage of 25(OH)D from mother to fetus could reduce maternal levels, especially if the mother is deficient in vitamin D; observational studies have shown either no change or a modest decline in maternal 25(OH)D concentrations during pregnancy (12, 13). No studies have addressed whether the ideal level of 25(OH)D during pregnancy should differ from the level considered sufficient for nonpregnant adults.

1,25(OH)₂D does not readily cross the placentas of rats (14), and 1,25(OH)₂D concentrations are normally lower than maternal values in fetal sheep, rats, mice, and humans (10, 11, 15, 16). The low fetal concentrations of 1,25(OH)₂D reflect the low fetal PTH and high phosphorus concentrations, which suppress renal 1 α -hydroxylase. Although PTHrP is elevated in the fetal circulation, it appears to be less able to stimulate the renal 1 α -hydroxylase than PTH (17, 18).

Total (free and bound) 1,25(OH)₂D concentrations double or triple in the maternal circulation starting in the first trimester, but studies have only shown increased free concentrations during the third trimester. This increase is due to maternal synthesis by the renal 1 α -hydroxylase (19, 20). Some have suggested that the fetus and placenta contribute to the maternal rise, but this is not the case, as shown by both animal studies (reviewed in detail in reference 2) and a clinical case of an anephric woman who had

only a negligible increase in $1,25(\text{OH})_2\text{D}$ concentrations during pregnancy (21).

Some researchers have argued that the doubled $1,25(\text{OH})_2\text{D}$ concentrations explain the doubling of intestinal calcium absorption and indicate the maternal adaptation's dependence on vitamin D sufficiency, but this explanation might be incomplete. Intestinal calcium absorption doubles in humans and rodents early in pregnancy, well before free $1,25(\text{OH})_2\text{D}$ concentrations increase late in pregnancy (2, 7). Furthermore, pregnant vitamin D–deficient rats and mice lacking the vitamin D receptor (VDR null) experience a marked up-regulation of intestinal calcium absorption to the same high rate as normal pregnant rats and mice (22–24). Animal studies indicate that prolactin and placental lactogen might stimulate intestinal calcium absorption independently of $1,25(\text{OH})_2\text{D}$ (25, 26). Although the data show that pregnant animals do not require $1,25(\text{OH})_2\text{D}$ and its receptor for intestinal calcium absorption doubling during pregnancy, no clinical study has compared intestinal calcium absorption during pregnancy in vitamin D–deficient and sufficient women.

VITAMIN D AND MATERNAL OUTCOMES FROM PREGNANCY

Animal data

Animal models used to examine vitamin D physiology during pregnancy and fetal development include severe vitamin D deficiency in rats (27–29), a naturally occurring null mutation of the 1α -hydroxylase in pigs (30), and VDR ablation in mice (24, 31). The 1α -hydroxylase has also been ablated in mice, but the null mice are infertile (32, 33).

In each of these models, the adult female has hypocalcemia, hypophosphatemia, rickets or osteomalacia, and reduced fertility with smaller litters. When such rats and mice do conceive, a few sporadic (possibly hypocalcemic) maternal deaths occur late in pregnancy during the interval of rapid calcium transfer to the fetus. Investigators have observed deaths in late pregnancy when giving rodents a low-calcium diet, which probably indicates that mothers rely on vitamin D and dietary calcium sufficiency to maintain their own blood calcium during ongoing rapid losses to multiple fetuses. In addition, vitamin D–deficient rats and VDR-null mice increased their skeletal mineral content during pregnancy (24, 29), although 1 study in vitamin D–deficient rats showed a small loss of skeletal mineral content during pregnancy (34). Studies in vitamin D–deficient rats and VDR-null mice have also shown that intestinal calcium absorption is upregulated to the normal pregnant level despite the respective absence of $1,25(\text{OH})_2\text{D}$ or its receptor.

Human data

No studies have focused specifically on vitamin D deficiency during pregnancy; the available data come from observational studies (12, 13, 35–41) and a few clinical trials of vitamin D supplementation (42–50) in pregnant women ranging from vitamin D deficient to sufficient. Severe vitamin D deficiency causes modest hypocalcemia and secondary hyperparathyroidism in nonpregnant adults, but no reports have documented worsening during pregnancy. Collectively, serum calcium concentrations were normal in women ranging from vitamin D deficient to sufficient. Many observational and randomized trials of pregnant women consistently showed that daily or monthly vitamin D_2 or

D_3 supplementation regimens can increase maternal $25(\text{OH})\text{D}$ concentrations, but none has shown any maternal benefit from such supplementation beyond the increase in circulating $25(\text{OH})\text{D}$. If the animal data apply to humans, they suggest that intestinal calcium absorption increases during pregnancy in women with severe vitamin D deficiency.

VITAMIN D AND FETAL OUTCOMES

Animal data

Studies of severely vitamin D–deficient rats (28, 51, 52), 1α -hydroxylase-deficient pigs (30), and VDR-null mice (31) have consistently shown strikingly normal fetal blood calcium, phosphorus, and PTH concentrations; fetal weight; and skeletal mineral and calcium content. 1α -Hydroxylase-null mice are normal at birth, but the literature includes no extensive studies of their fetal chemistries and skeletal mineral content (32, 33). Researchers have assayed placental calcium transfer from mother to fetus indirectly in vitamin D–deficient rats (53) and directly in VDR-null fetuses (31); calcium concentrations were normal to nonsignificantly increased in both. Clearly, fetal calcium homeostasis and skeletal development and mineralization are independent of vitamin D, $1,25(\text{OH})_2\text{D}$, and its receptor. The placenta provides calcium without relying on vitamin D metabolites, and vitamin D–deficient and VDR-null placentas express normal concentrations of the vitamin D–dependent factors calbindin-D-9K and Ca^{2+} -ATPase, which are important for intestinal calcium absorption and calcium homeostasis in adults (31, 53, 54).

Offspring of VDR-heterozygous mice (wild-type, heterozygous-deleted, and VDR-null fetuses) were indistinguishable with respect to calcium homeostasis, weight, skeletal size, morphology, and mineral content. Fetuses of VDR-null mothers had a lower birth weight than did those with VDR-heterozygous mothers, but their proportionately smaller skeletons had a normal mineral content (31). Researchers did not observe this global reduction in fetal size and weight from VDR-null mothers in vitamin D deficiency models, which could indicate that absence of VDR affects fetal growth in a way that absence of vitamin D does not.

In animal models of maternal hypoparathyroidism, fetuses can develop secondary hyperparathyroidism, skeletal demineralization, and fractures (2). Researchers have not observed this with vitamin D deficiency, perhaps because the maternal calcium level is usually less low from vitamin D deficiency than from hypoparathyroidism.

Human data

No systematic studies have examined skeletal mineral content among normal, vitamin D–insufficient, and vitamin D–deficient fetuses; thus, we do not know whether vitamin D–deficient or –insufficient human fetuses have normal skeletal mineral content as studies have shown for vitamin D–deficient animals. Clinical experience reported in textbook chapters and reviews indicates that fetuses with severe vitamin D deficiency are generally born with normal serum calcium concentrations and skeletal morphology, and rickets does not develop (or clinicians do not recognize it) until weeks to months after birth (55–57). The observational and clinical studies of human pregnancy cited above showed no relation of cord blood $25(\text{OH})\text{D}$ concentrations to cord blood calcium or PTH concentrations.



Those observational studies and clinical trials showed that providing vitamin D to pregnant mothers increases cord 25(OH)D concentrations at term but has no effect on fetal weight or skeletal parameters. Several studies found no effect of maternal vitamin D supplementation on cord blood calcium concentrations (42, 48, 49), but 2 small studies in Asian women showed a small but significant increase in cord calcium concentrations (46, 47). Another study compared administration of 1200 mg Ca and 10 μg (400 IU) vitamin D (as dairy) or 1200 mg Ca alone (as orange juice) with placebo and found greater birth weight and total body calcium levels in the fetuses whose mothers received the dairy product but no change in other skeletal variables such as length and head circumference (50). The authors do not know whether the results were due to the dairy product's vitamin D content or its nutritional content compared with the supplement.

Studies of mother-infant pairs have shown no convincing relation between maternal vitamin D sufficiency and fetal outcomes at birth. One study found no association between third-trimester maternal 25(OH)D concentrations and any fetal measurement, but offspring of women with 25(OH)D concentrations <25 nmol/L during the third trimester had a knee-heel length 2.7 mm shorter (not statistically significant) after the authors corrected for gestational length (39). Another study found no association of third-trimester 25(OH)D concentrations with any fetal measurement, including weight, head circumference, arm circumference, and length (40, 41). No systematic study has investigated skeletal lengths and calcium content (by dual-energy X-ray absorptiometry) in newborns stratified by vitamin D status, but the available animal and human data indicate that vitamin D status should have little or no effect on the fetal skeleton's length and mineral content.

Overall, whereas animal studies have shown normal serum calcium concentrations, skeletal lengths, and skeletal mineral content in fetuses despite extreme disturbances in vitamin D physiology, none of the human studies has approached this level of careful, systematic investigation. Consequently, the possibility remains that the human studies lacked the power or sensitive outcome measures required to detect differences among normal, vitamin D–insufficient, and vitamin D–deficient fetuses.

ADAPTATIONS DURING LACTATION

Near-exclusive breastfeeding for 6 mo leads, on average, to maternal calcium loss 4 times higher than in pregnancy because lactation can require $150\text{--}300$ mg Ca \cdot kg⁻¹ \cdot d⁻¹. Characteristic findings (2, 3, 5) in the blood chemistries of healthy lactating women include that serum calcium and ionized calcium concentrations are normal, although some reports suggest that ionized or corrected serum calcium concentrations rise slightly but stay within the normal range. Phosphorus can rise above the normal range, probably because of accelerated resorption from the skeleton (discussed below). PTH concentrations, as measured by “intact” assays, fall to the lower end of the normal range or below, except in women known to have a low calcium or vitamin D intake, including women from Asia and Gambia. Estradiol concentrations are low and near menopausal values. Prolactin concentrations increase at each suckling, but the basal concentrations between feeds decline with time postpartum. PTHrP concentrations are higher than PTH concentrations in nonpregnant women and show some pulsatility in response to suckling.

The lactation-induced adaptations to maternal calcium homeostasis are illustrated in Figure 1. As described in detail elsewhere (5, 6, 8), PTHrP (produced by the lactating breast) in combination with low estradiol concentrations appears to drive the main physiologic adaptation to meet the calcium demands of lactation (Figure 2). Suckling and prolactin both inhibit ovarian function and stimulate PTHrP. Together, PTHrP and low estradiol concentrations stimulate skeletal resorption, and bone mineral content declines by 5–10% over 2–6 mo of near-exclusive lactation. Bone resorption markers show marked elevation without a compensatory increase in bone formation. Intestinal calcium absorption rates drop to the normal range after delivery. Renal calcium reabsorption rates increase, presumably due to PTHrP, which mimics the actions of PTH on the renal tubules.

Lactating rodents have a similar adaptive mechanism and lose 25–35% of their trabecular bone mineral content during 3 wk of lactation. Similarly, PTH concentrations are usually low but increase with litter size, estradiol concentrations are low, and PTHrP concentrations are high. Intestinal calcium absorption rates are still approximately double those of nonpregnant animals (22, 58), which might be necessary to meet the proportionately greater calcium demands of multiple suckling pups. Animal models also show that local actions of the calcium receptor and PTHrP within mammary tissue regulate milk's calcium content, at least partly.

VITAMIN D METABOLISM DURING LACTATION

Vitamin D passes readily into breast milk, 25(OH)D passes very poorly, and 1,25(OH)₂D does not appear to pass at all (2). 1,25(OH)₂D concentrations fall rapidly after pregnancy and are normal during lactation, except in women nursing twins, who have increased 1,25(OH)₂D concentrations (2). 25(OH)D concentrations were stable in 1 study (59) but fell during lactation in another (60). Breast milk should only account for a small loss of 25(OH)D; seasonal variation and differences in vitamin supplement use before and after pregnancy might have confounded the results. In lactating rats and mice, 1,25(OH)₂D concentrations remain elevated until weaning (2).

VITAMIN D AND MATERNAL OUTCOMES FROM LACTATION

Animal data

Not only are 1,25(OH)₂D concentrations elevated during lactation in normal rodents, but the concentrations respond to varying lactation demands. When stressed by a low-calcium diet or large litter size, 1,25(OH)₂D concentrations increase even further (61, 62), perhaps because a mechanism increases intestinal calcium absorption further when the mother faces extra demands. However, mothers do not require vitamin D sufficiency or responsiveness to 1,25(OH)₂D for normal lactation. Vitamin D–deficient rats and VDR-null mice lactate normally and experience similar skeletal losses to controls (24, 29, 34), although 1 study found that vitamin D–deficient rats lose more skeletal mineral content than do normal rats (63). Intestinal calcium absorption in lactating vitamin D–deficient rats is upregulated to the same level as in vitamin D–sufficient rats (22, 58).

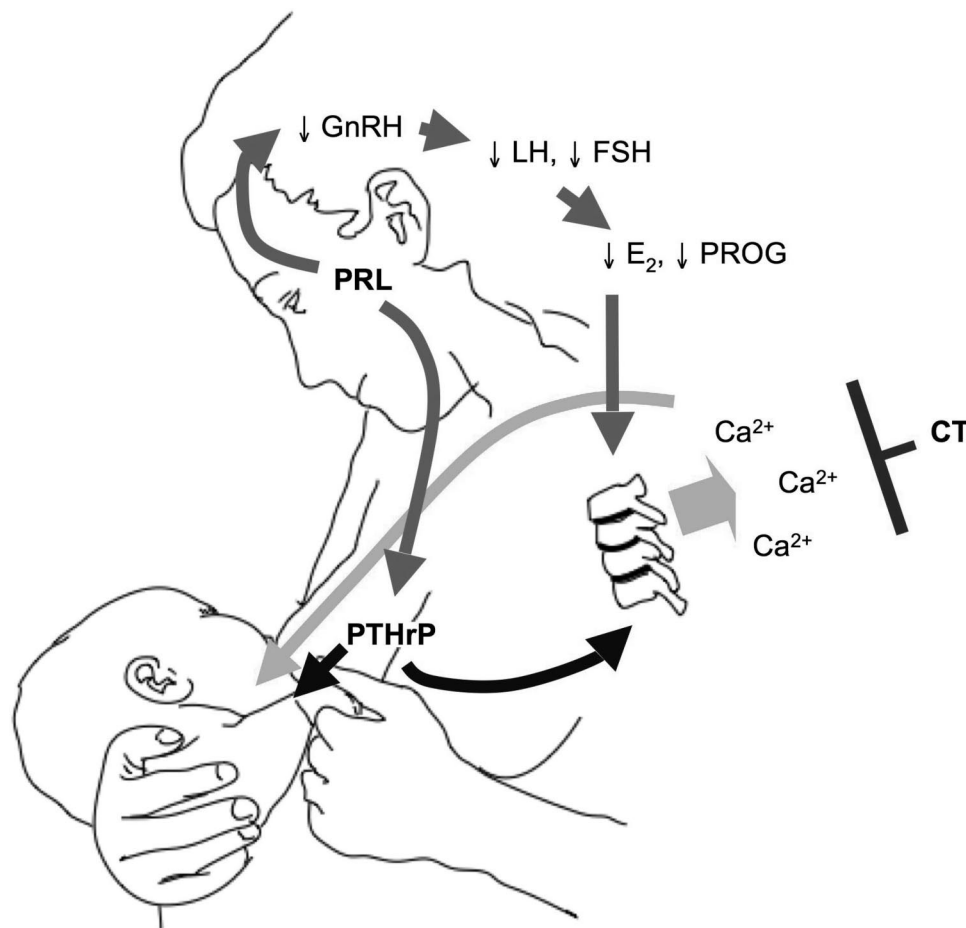


FIGURE 2. The breast is a central regulator of skeletal demineralization during lactation. Suckling induces prolactin (PRL) release. Suckling and prolactin both inhibit the hypothalamic gonadotropin-releasing hormone (GnRH) pulse center, which in turn suppresses the gonadotropins [luteinizing hormone (LH) and follicle-stimulating hormone (FSH)], leading to low levels of the ovarian sex steroids [estradiol (E₂) and progesterone (PROG)]. Several factors control parathyroid hormone-related protein (PTHrP) production and release from the breast, including suckling, prolactin, and the calcium receptor. PTHrP enters the bloodstream and combines with systemically low estradiol concentrations to markedly up-regulate bone resorption. Increased bone resorption releases calcium and phosphate into the bloodstream, which then reaches the breast ducts and is actively pumped into the breast milk. PTHrP also passes into milk at high concentrations, but we do not know whether swallowed PTHrP plays a role in regulating the neonate's calcium physiology. Calcitonin (CT) might inhibit skeletal responsiveness to PTHrP and low estradiol. Reprinted from reference 8, copyright 2005, with kind permission of Springer Science and Business Media B.

Human data

Observational studies (59, 60, 64–67) and clinical trials (68–75) have generally shown that providing vitamin D to lactating mothers increases their 25(OH)D concentrations but has no significant effect on any other maternal outcome (68, 69, 72–74). Randomized trials and observational studies of dietary calcium intake's effect on skeletal resorption during lactation in North American and Gambian women have consistently shown that very low to well-above-normal calcium intakes had no effect on skeletal demineralization during lactation but did increase urinary calcium excretion (76–81). Most of these studies compared calcium intake and did not manipulate vitamin D intake directly, but a test of vitamin D supplementation during lactation would probably find no effect on skeletal resorption. Limited studies of lactating adolescents have reported greater skeletal losses than in older women, perhaps due to poor calcium intake and nutrition in the adolescents (82, 83). In other studies, maternal vitamin D status or vitamin D supplementation did not affect breast milk calcium content (73, 84).

Thus, while one might expect that low vitamin D and calcium intakes would accentuate skeletal losses to maintain breast milk calcium content, most studies suggest otherwise. This is consistent with the animal studies and might indicate that skeletal resorption provides most of the calcium needed during lactation, regardless of dietary calcium intake. The obligatory rise in PTHrP and fall in estradiol programs the lactational loss of skeletal calcium content (Figure 2), and vitamin D status does not influence this loss. Increasing calcium and vitamin D intake during lactation might simply increase urinary calcium excretion and, thereby, kidney stone risk.

VITAMIN D AND NEONATAL AND INFANT OUTCOMES

Animal data

In vitamin D–deficient rats (51, 52), 1 α -hydroxylase-null pigs (30), VDR-null mice (85, 86), and 1 α -hydroxylase-null mice



(32, 33), rickets and failure to thrive are not apparent until near weaning. Studies of vitamin D–deficient rats and VDR-null mice confirm that skeletal mineral content is normal at birth and during the first 2 wk after birth, after which the animals develop progressive hypocalcemia, hypophosphatemia, and histomorphometric evidence of rickets. The sequence of events parallels the maturation of intestinal calcium absorption postnatally, which develops from nonsaturable passive absorption facilitated by lactose to an active, saturable process that depends on $1,25(\text{OH})_2\text{D}$ (87–89).

Human data

No systematic studies have addressed the effects of vitamin D status on neonatal or infant calcium and bone status parameters; case reports and clinical experiences described in textbook chapters constitute the main data. Vitamin D deficiency predisposes newborns to neonatal hypocalcemia, and clinicians do not usually diagnose (or recognize) rickets for several months after birth (55–57). However, in areas where vitamin D deficiency is endemic and clinical awareness is high, clinicians often identify the characteristic changes of rickets soon after birth (90, 91).

After birth, serum calcium concentrations drop from their high fetal levels to a trough below the adult level, followed by a gradual recovery over several days to the adult level (2, 92). Although vitamin D deficiency increases neonatal hypocalcemia risk, it is unclear whether vitamin D insufficiency causes hypocalcemia. Vitamin D supplementation during pregnancy reduced blood calcium excursion in neonates in 1 study and reduced hypocalcemia incidence in another (42, 48, 49).

Standard 10 μg (400 IU) vitamin D supplements given to lactating mothers do not increase infant $25(\text{OH})\text{D}$ concentrations because of $25(\text{OH})\text{D}$'s poor penetrance into milk; a dose of 100 μg (4000 IU) per day was required to raise neonatal $25(\text{OH})\text{D}$ concentrations to the perceived sufficient range of >75 nmol/L (72). Dosing the infant directly with smaller doses of vitamin D produces normal $25(\text{OH})\text{D}$ concentrations but vitamin D supplementation in otherwise healthy infants (via mother's milk or directly to the infant) did not improve the infants' blood calcium concentrations, length, weight, or other parameters.

One study found no association between third-trimester maternal $25(\text{OH})\text{D}$ concentrations and newborn weight, length, or head circumference indexes (40). Follow-up assessments of these children also found no effect at 9 mo or 9 y of age (40, 41). However, bone mineral content was apparently lower in 9-y-old children whose mothers had had low $25(\text{OH})\text{D}$ (<20 nmol/L) concentrations during the third trimester (40), which might indicate the effect of in utero vitamin D status to program peak bone mass that will be achieved later in life (93, 94). Because the investigators did not measure bone mineral content at birth or 9 mo, it is not clear whether this is truly a mechanistic association [low fetal $25(\text{OH})\text{D}$ programming low bone mineral content] or is related to environment and nutrition [because a pregnant woman with a low $25(\text{OH})\text{D}$ level might be more likely to provide poor nutrition to her child, have lower socioeconomic status, etc].

Observational studies have shown that up to 95% of children with vitamin D–deficient rickets had been breastfed (95), which is consistent with the milk's low vitamin D and $25(\text{OH})\text{D}$ contents unless the woman takes supplements aggressively. Supplementing the mother during pregnancy to provide the infant with normal $25(\text{OH})\text{D}$ stores at birth or supplementing the infant directly can prevent childhood rickets.

Increasing evidence from observational studies indicates that vitamin D deficiency and insufficiency at older ages might increase the risk of chronic diseases such as type 1 diabetes and multiple sclerosis. In many of these diseases, the association is with latitude and, by inference, with vitamin D status. For most of these associations, no specific data relate the disease to fetal or neonatal vitamin D sufficiency. However, observational studies indicate that vitamin D insufficiency during pregnancy is associated with increased prevalence of islet cell antibodies in offspring, and a history of vitamin D supplementation in pregnant women (96, 97) or infants (98) is associated with lower childhood incidence of type 1 diabetes. Investigators need to conduct randomized trials on this association before clinicians recommend vitamin D to reduce the incidence of type 1 diabetes. Although human fetuses might suffer no skeletal problems from vitamin D deficiency and insufficiency, they could have an increased risk of nonskeletal problems, such as type 1 diabetes, in childhood.

POSTWEANING SKELETAL RECOVERY

After weaning, the maternal skeleton rapidly recovers the mineral content lost during lactation. In clinical studies, the recovery apparently occurred within 3–6 mo, although many studies did not follow the women after weaning. Observational studies generally indicate that a history of lactation confers no increased risk of low bone mass, fractures, or osteoporosis (2, 99).

This recovery is especially remarkable when one considers that the adult skeleton normally recovers incompletely, if at all, from bone mass losses induced by weightlessness, bed rest, corticosteroid therapy, estrogen deficiency, etc. We do not know what mechanism explains skeletal recovery after lactation. One observational study showed that intestinal calcium absorption was upregulated by 19% during postweaning recovery (71), but the investigators assayed only 1 time point and the magnitude was quite modest.

Lactating rodents recover completely within 10–21 d, depending on the rodent strain and technique used. No studies have systematically measured intestinal calcium absorption or calcitropic hormone concentrations during the recovery interval.

Animal data

Two studies of vitamin D–deficient rats noted some recovery of mineral content after lactation, with the final value exceeding the prepregnancy value in 1 study (29, 34). In preliminary studies of VDR-null mice, skeletal recovery after lactation was complete and final bone mineral content exceeded the prepregnancy level by 50% (24). Thus, animal studies suggest that vitamin D status plays no role in skeletal recovery after lactation.

Human data

No study has examined the impact of vitamin D deficiency or insufficiency on the skeleton's ability to recover from lactational losses. One study of lactating women observed that PTH and $1,25(\text{OH})_2\text{D}$ concentrations were higher than normal at 1 time point assayed during postweaning recovery (100). No other study has examined this, so we do not know whether the skeleton requires vitamin D sufficiency to recover. Vitamin D requirements might be higher during postweaning recovery than after, but this is also speculation.

CONCLUSIONS

Vitamin D deficiency during pregnancy and lactation can lead to hypocalcemia and rickets in neonates and, especially, infants, but animal data and limited human data suggest that fetuses are protected from the adverse skeletal effects of vitamin D deficiency. Adaptations in maternal calcium and bone metabolism appear to occur independently of vitamin D status. Careful study of newborns with vitamin D-deficient mothers might reveal deficits in skeletal mineral content by dual-energy X-ray absorptiometry. Given the apparent relative protection of mothers and fetuses from severe vitamin D deficiency, vitamin D insufficiency probably does not harm the fetus, infant, or mother. Dosing recommendations for mothers during pregnancy should be aimed at preventing problems in neonates and infants, and a vitamin D dose sufficient for the mother during pregnancy should produce normal cord blood 25(OH)D concentrations at birth. Giving relatively small doses of vitamin D directly to the infant or supplementing the mother with 100 μ g (4000 IU) vitamin D daily should maintain normal 25(OH)D concentrations in exclusively breastfed infants without harming the mother. Researchers need to study aspects of the role of vitamin D sufficiency and supplementation in pregnancy and lactation, especially the relation (if any) between vitamin D insufficiency in utero and infancy and long-term outcomes such as type 1 diabetes, multiple sclerosis, and other chronic diseases.

The author had no conflict of interest.

REFERENCES

- Dawson-Hughes B. Calcium and vitamin D. In: Favus MJ, ed. *Primer on the metabolic bone diseases and disorders of mineral metabolism*. 6th ed. Washington, DC: ASBMR Press, 2006;257-9.
- Kovacs CS, Kronenberg HM. Maternal-fetal calcium and bone metabolism during pregnancy, puerperium and lactation. *Endocr Rev* 1997; 18:832-72.
- Kovacs CS, El-Hajj Fuleihan G. Calcium and bone disorders during pregnancy and lactation. *Endocrinol Metab Clin N Am* 2006;35:21-51.
- Kovacs CS. Fetal mineral homeostasis. In: Glorieux FH, Pettifor JM, Jüppner H, eds. *Pediatric bone: biology and diseases*. San Diego, CA: Academic Press, 2003;271-302.
- Wysolmerski JJ. Conversations between breast and bone: physiological bone loss during lactation as evolutionary template for osteolysis in breast cancer and pathological bone loss after menopause. *Bonekey Osteovision* 2007;4:209-25.
- Wysolmerski JJ. The evolutionary origins of maternal calcium and bone metabolism during lactation. *J Mammary Gland Biol Neoplasia* 2002;7:267-76.
- Kovacs CS. Calcium and bone metabolism in pregnancy and lactation. *J Clin Endocrinol Metab* 2001;86:2344-8.
- Kovacs CS. Calcium and bone metabolism during pregnancy and lactation. *J Mammary Gland Biol Neoplasia* 2005;10:105-18.
- Haddad JG Jr, Boisseau V, Avioli LV. Placental transfer of vitamin D₃ and 25-hydroxycholecalciferol in the rat. *J Lab Clin Med* 1971;77:908-15.
- Fleischman AR, Rosen JF, Cole J, Smith CM, DeLuca HF. Maternal and fetal serum 1,25-dihydroxyvitamin D levels at term. *J Pediatr* 1980;97:640-2.
- Wieland P, Fischer JA, Trechsel U, et al. Perinatal parathyroid hormone, vitamin D metabolites, and calcitonin in man. *Am J Physiol* 1980;239:E385-90.
- Hillman LS, Slatopolsky E, Haddad JG. Perinatal vitamin D metabolism. IV. Maternal and cord serum 24,25-dihydroxyvitamin D concentrations. *J Clin Endocrinol Metab* 1978;47:1073-7.
- Ardawi MS, Nasrat HA, BA'Aqueel HS. Calcium-regulating hormones and parathyroid hormone-related peptide in normal human pregnancy and postpartum: a longitudinal study. *Eur J Endocrinol* 1997; 137:402-9.
- Noff D, Edelstein S. Vitamin D and its hydroxylated metabolites in the rat. Placental and lacteal transport, subsequent metabolic pathways and tissue distribution. *Horm Res* 1978;9:292-300.
- Seki K, Furuya K, Makimura N, Mitsui C, Hirata J, Nagata I. Cord blood levels of calcium-regulating hormones and osteocalcin in premature infants. *J Perinat Med* 1994;22:189-94.
- Hollis BW, Pittard WB. Evaluation of the total fetomaternal vitamin D relationships at term: evidence for racial differences. *J Clin Endocrinol Metab* 1984;59:652-7.
- Horwitz MJ, Tedesco MB, Sereika SM, Hollis BW, Garcia-Ocana A, Stewart AF. Direct comparison of sustained infusion of human parathyroid hormone-related protein-(1-36). *J Clin Endocrinol Metab* 2003; 88:1603-9.
- Horwitz MJ, Tedesco MB, Sereika SM, et al. Continuous PTH and PTHrP infusion causes suppression of bone formation and discordant effects on 1,25(OH)₂ vitamin D. *J Bone Miner Res* 2005;20:1792-803.
- Fenton E, Britton HG. 25-Hydroxycholecalciferol 1 alpha-hydroxylase activity in the kidney of the fetal, neonatal and adult guinea pig. *Biol Neonate* 1980;37:254-6.
- Kubota M, Ohno J, Shiina Y, Suda T. Vitamin D metabolism in pregnant rabbits: differences between the maternal and fetal response to administration of large amounts of vitamin D₃. *Endocrinology* 1982; 110:1950-6.
- Turner M, Barre PE, Benjamin A, Goltzman D, Gascon-Barre M. Does the maternal kidney contribute to the increased circulating 1,25-dihydroxyvitamin D concentrations during pregnancy? *Miner Electrolyte Metab* 1988;14:246-52.
- Halloran BP, DeLuca HF. Calcium transport in small intestine during pregnancy and lactation. *Am J Physiol* 1980;239:E64-8.
- Brommage R, Baxter DC, Gierke LW. Vitamin D-independent intestinal calcium and phosphorus absorption during reproduction. *Am J Physiol* 1990;259:G631-8.
- Fudge NJ, Woodrow JP, Kovacs CS. Pregnancy rescues low bone mass and normalizes intestinal calcium absorption in Vdr null mice. *J Bone Miner Res* 2006;21(Suppl):S52.
- Pahuja DN, DeLuca HF. Stimulation of intestinal calcium transport and bone calcium mobilization by prolactin in vitamin D-deficient rats. *Science* 1981;214:1038-9.
- Mainoya JR. Effects of bovine growth hormone, human placental lactogen and ovine prolactin on intestinal fluid and ion transport in the rat. *Endocrinology* 1975;96:1165-70.
- Halloran BP, Barthell EN, DeLuca HF. Vitamin D metabolism during pregnancy and lactation in the rat. *Proc Natl Acad Sci U S A* 1979;76: 5549-53.
- Halloran BP, DeLuca HF. Vitamin D deficiency and reproduction in rats. *Science* 1979;204:73-4.
- Halloran BP, DeLuca HF. Skeletal changes during pregnancy and lactation: the role of vitamin D. *Endocrinology* 1980;107:1923-9.
- Lachenmaier-Currle U, Harmeyer J. Placental transport of calcium and phosphorus in pigs. *J Perinat Med* 1989;17:127-36.
- Kovacs CS, Woodland ML, Fudge NJ, Friel JK. The vitamin D receptor is not required for fetal mineral homeostasis or for the regulation of placental calcium transfer. *Am J Physiol Endocrinol Metab* 2005;289: E133-44.
- Panda DK, Miao D, Tremblay ML, et al. Targeted ablation of the 25-hydroxyvitamin D 1alpha-hydroxylase enzyme: evidence for skeletal, reproductive, and immune dysfunction. *Proc Natl Acad Sci U S A* 2001;98:7498-503.
- Dardenne O, Prud'homme J, Arabian A, Glorieux FH, St-Arnaud R. Targeted inactivation of the 25-hydroxyvitamin D(3)-1(alpha)-hydroxylase gene (CYP27B1) creates an animal model of pseudovitamin D-deficiency rickets. *Endocrinology* 2001;142:3135-41.
- Miller SC, Halloran BP, DeLuca HF, Jee WS. Role of vitamin D in maternal skeletal changes during pregnancy and lactation: a histomorphometric study. *Calcif Tissue Int* 1982;34:245-52.
- Paunier L, Lacourt G, Pilloud P, Schlaeppli P, Sizonenko PC. 25-Hydroxyvitamin D and calcium levels in maternal, cord and infant serum in relation to maternal vitamin D intake. *Helv Paediatr Acta* 1978;33:95-103.
- Congdon P, Horsman A, Kirby PA, Dibble J, Bashir T. Mineral content of the forearms of babies born to Asian and white mothers. *Br Med J (Clin Res Ed)* 1983;286:1233-5.
- Madelenat P, Bastian H, Menn S. [Winter supplementation in the 3rd



- trimester of pregnancy by a dose of 80,000 IU of vitamin D]. *J Gynecol Obstet Biol Reprod (Paris)* 2001;30:761–7 (in French).
38. Datta S, Alfaham M, Davies DP, et al. Vitamin D deficiency in pregnant women from a non-European ethnic minority population—an interventional study. *BJOG* 2002;109:905–8.
 39. Morley R, Carlin JB, Pasco JA, Wark JD. Maternal 25-hydroxyvitamin D and parathyroid hormone concentrations and offspring birth size. *J Clin Endocrinol Metab* 2006;91:906–12.
 40. Javaid MK, Crozier SR, Harvey NC, et al. Maternal vitamin D status during pregnancy and childhood bone mass at age 9 years: a longitudinal study. *Lancet* 2006;367:36–43.
 41. Gale CR, Robinson SM, Harvey NC, et al. Maternal vitamin D status during pregnancy and child outcomes. *Eur J Clin Nutr* 2008;62:68–77.
 42. Brooke OG, Brown IR, Bone CD, et al. Vitamin D supplements in pregnant Asian women: effects on calcium status and fetal growth. *Br Med J* 1980;280:751–4.
 43. Brooke OG, Butters F, Wood C. Intrauterine vitamin D nutrition and postnatal growth in Asian infants. *Br Med J (Clin Res Ed)* 1981;283:1024.
 44. Maxwell JD, Ang L, Brooke OG, Brown IR. Vitamin D supplements enhance weight gain and nutritional status in pregnant Asians. *Br J Obstet Gynaecol* 1981;88:987–91.
 45. Cockburn F, Belton NR, Purvis RJ, et al. Maternal vitamin D intake and mineral metabolism in mothers and their newborn infants. *Br Med J* 1980;281:11–4.
 46. Marya RK, Rathee S, Dua V, Sangwan K. Effect of vitamin D supplementation during pregnancy on foetal growth. *Indian J Med Res* 1988;88:488–92.
 47. Marya RK, Rathee S, Lata V, Mudgil S. Effects of vitamin D supplementation in pregnancy. *Gynecol Obstet Invest* 1981;12:155–61.
 48. Delvin EE, Salle BL, Glorieux FH, Adeleine P, David LS. Vitamin D supplementation during pregnancy: effect on neonatal calcium homeostasis. *J Pediatr* 1986;109:328–34.
 49. Mallet E, Gugi B, Brunelle P, Henocq A, Basuyau JP, Lemeur H. Vitamin D supplementation in pregnancy: a controlled trial of two methods. *Obstet Gynecol* 1986;68:300–4.
 50. Chan GM, McElligott K, McNaught T, Gill G. Effects of dietary calcium intervention on adolescent mothers and newborns: a randomized controlled trial. *Obstet Gynecol* 2006;108:565–71.
 51. Halloran BP, De Luca HF. Effect of vitamin D deficiency on skeletal development during early growth in the rat. *Arch Biochem Biophys* 1981;209:7–14.
 52. Miller SC, Halloran BP, DeLuca HF, Jee WS. Studies on the role of vitamin D in early skeletal development, mineralization, and growth in rats. *Calcif Tissue Int* 1983;35:455–60.
 53. Glazier JD, Mawer EB, Sibley CP. Calbindin-D_{9k} gene expression in rat chorioallantoic placenta is not regulated by 1,25-dihydroxyvitamin D₃. *Pediatr Res* 1995;37:720–5.
 54. Marche P, Delorme A, Cuisinier-Gleizes P. Intestinal and placental calcium-binding proteins in vitamin D-deprived or -supplemented rats. *Life Sci* 1978;23:2555–61.
 55. Campbell DE, Fleischman AR. Rickets of prematurity: controversies in causation and prevention. *Clin Perinatol* 1988;15:879–90.
 56. Pereira GR, Zucker AH. Nutritional deficiencies in the neonate. *Clin Perinatol* 1986;13:175–89.
 57. Specker BL. Do North American women need supplemental vitamin D during pregnancy or lactation? *Am J Clin Nutr* 1994;59(suppl):484S–90S.
 58. Boass A, Toverud SU, Pike JW, Haussler MR. Calcium metabolism during lactation: enhanced intestinal calcium absorption in vitamin D-deprived, hypocalcemic rats. *Endocrinology* 1981;109:900–7.
 59. Kent GN, Price RI, Gutteridge DH, et al. Human lactation: forearm trabecular bone loss, increased bone turnover, and renal conservation of calcium and inorganic phosphate with recovery of bone mass following weaning. *J Bone Miner Res* 1990;5:361–9.
 60. Sowers M, Zhang D, Hollis BW, et al. Role of calciotropic hormones in calcium mobilization of lactation. *Am J Clin Nutr* 1998;67:284–91.
 61. Lobaugh B, Boass A, Garner SC, Toverud SU. Intensity of lactation modulates renal 1 α -hydroxylase and serum 1,25(OH)₂D in rats. *Am J Physiol* 1992;262:E840–4.
 62. Lobaugh B, Boass A, Lester GE, Toverud SU. Regulation of serum 1,25-dihydroxyvitamin D₃ in lactating rats. *Am J Physiol* 1990;259:E665–71.
 63. Marie PJ, Cancela L, Le Boulch N, Miravet L. Bone changes due to pregnancy and lactation: influence of vitamin D status. *Am J Physiol* 1986;251:E400–6.
 64. Cancela L, Le Boulch N, Miravet L. Relationship between the vitamin D content of maternal milk and the vitamin D status of nursing women and breast-fed infants. *J Endocrinol* 1986;110:43–50.
 65. Okonofua F, Menon RK, Houlder S, et al. Calcium, vitamin D and parathyroid hormone relationships in pregnant Caucasian and Asian women and their neonates. *Ann Clin Biochem* 1987;24:22–8.
 66. Takeuchi A, Okano T, Tsugawa N, et al. Effects of ergocalciferol supplementation on the concentration of vitamin D and its metabolites in human milk. *J Nutr* 1989;119:1639–46.
 67. Alfaham M, Woodhead S, Pask G, Davies D. Vitamin D deficiency: a concern in pregnant Asian women. *Br J Nutr* 1995;73:881–7.
 68. Rothberg AD, Pettifor JM, Cohen DF, Sonnendecker EW, Ross FP. Maternal-infant vitamin D relationships during breast-feeding. *J Pediatr* 1982;101:500–3.
 69. Ala-Houhala M. 25-Hydroxyvitamin D levels during breast-feeding with or without maternal or infantile supplementation of vitamin D. *J Pediatr Gastroenterol Nutr* 1985;4:220–6.
 70. Ala-Houhala M, Koskinen T, Parviainen MT, Visakorpi JK. 25-Hydroxyvitamin D and vitamin D in human milk: effects of supplementation and season. *Am J Clin Nutr* 1988;48:1057–60.
 71. Kalkwarf HJ, Specker BL, Heubi JE, Vieira NE, Yergey AL. Intestinal calcium absorption of women during lactation and after weaning. *Am J Clin Nutr* 1996;63:526–31.
 72. Hollis BW, Wagner CL. Vitamin D requirements during lactation: high-dose maternal supplementation as therapy to prevent hypovitaminosis D for both the mother and the nursing infant. *Am J Clin Nutr* 2004;80(suppl):1752S–8S.
 73. Basile LA, Taylor SN, Wagner CL, Horst RL, Hollis BW. The effect of high-dose vitamin D supplementation on serum vitamin D levels and milk calcium concentration in lactating women and their infants. *Breastfeed Med* 2006;1:27–35.
 74. Wagner CL, Hulsey TC, Fanning D, Ebeling M, Hollis BW. High-dose vitamin D₃ supplementation in a cohort of breastfeeding mothers and their infants: a 6-month follow-up pilot study. *Breastfeed Med* 2006;1:59–70.
 75. Saadi HF, Dawodu A, Afandi BO, Zayed R, Benedict S, Nagelkerke N. Efficacy of daily and monthly high-dose calciferol in vitamin D-deficient nulliparous and lactating women. *Am J Clin Nutr* 2007;85:1565–71.
 76. Cross NA, Hillman LS, Allen SH, Krause GF. Changes in bone mineral density and markers of bone remodeling during lactation and postweaning in women consuming high amounts of calcium. *J Bone Miner Res* 1995;10:1312–20.
 77. Kalkwarf HJ, Specker BL, Bianchi DC, Ranz J, Ho M. The effect of calcium supplementation on bone density during lactation and after weaning. *N Engl J Med* 1997;337:523–8.
 78. Polatti F, Capuzzo E, Viazzo F, Colleoni R, Klersy C. Bone mineral changes during and after lactation. *Obstet Gynecol* 1999;94:52–6.
 79. Laskey MA, Prentice A, Hanratty LA, et al. Bone changes after 3 mo of lactation: influence of calcium intake, breast-milk output, and vitamin D-receptor genotype. *Am J Clin Nutr* 1998;67:685–92.
 80. Fairweather-Tait S, Prentice A, Heumann KG, et al. Effect of calcium supplements and stage of lactation on the calcium absorption efficiency of lactating women accustomed to low calcium intakes. *Am J Clin Nutr* 1995;62:1188–92.
 81. Prentice A, Jarjou LM, Cole TJ, Stirling DM, Dibba B, Fairweather-Tait S. Calcium requirements of lactating Gambian mothers: effects of a calcium supplement on breast-milk calcium concentration, maternal bone mineral content, and urinary calcium excretion. *Am J Clin Nutr* 1995;62:58–67.
 82. Chan GM, Ronald N, Slater P, Hollis J, Thomas MR. Decreased bone mineral status in lactating adolescent mothers. *J Pediatr* 1982;101:767–70.
 83. Chan GM, McMurry M, Westover K, Engelbert-Fenton K, Thomas MR. Effects of increased dietary calcium intake upon the calcium and bone mineral status of lactating adolescent and adult women. *Am J Clin Nutr* 1987;46:319–23.
 84. Prentice A, Yan L, Jarjou LM, et al. Vitamin D status does not influence the breast-milk calcium concentration of lactating mothers accustomed to a low calcium intake. *Acta Paediatr* 1997;86:1006–8.
 85. Li YC, Amling M, Pirro AE, et al. Normalization of mineral ion homeostasis by dietary means prevents hyperparathyroidism, rickets, and

- osteomalacia, but not alopecia in vitamin D receptor-ablated mice. *Endocrinology* 1998;139:4391–6.
86. Li YC, Pirro AE, Amling M, et al. Targeted ablation of the vitamin D receptor: an animal model of vitamin D dependent rickets type II with alopecia. *Proc Natl Acad Sci U S A* 1997;94:9831–5.
87. Halloran BP, DeLuca HF. Calcium transport in small intestine during early development: role of vitamin D. *Am J Physiol* 1980;239:G473–9.
88. Ghishan FK, Parker P, Nichols S, Hoyumpa A. Kinetics of intestinal calcium transport during maturation in rats. *Pediatr Res* 1984;18:235–9.
89. Ghishan FK, Jenkins JT, Younoszai MK. Maturation of calcium transport in the rat small and large intestine. *J Nutr* 1980;110:1622–8.
90. Teotia M, Teotia SP. Nutritional and metabolic rickets. *Indian J Pediatr* 1997;64:153–7.
91. Teotia M, Teotia SP, Nath M. Metabolic studies in congenital vitamin D deficiency rickets. *Indian J Pediatr* 1995;62:55–61.
92. Kovacs CS. Skeletal physiology: fetus and neonate. In: Favus MJ, ed. *Primer on the metabolic bones diseases and disorders of mineral metabolism*. 6th ed. Washington, DC: ASBMR Press, 2006;50–5.
93. Cooper C, Javaid K, Westlake S, Harvey N, Dennison E. Developmental origins of osteoporotic fracture: the role of maternal vitamin D insufficiency. *J Nutr* 2005;135:2728S–34S.
94. Cooper C, Westlake S, Harvey N, Javaid K, Dennison E, Hanson M. Review: developmental origins of osteoporotic fracture. *Osteoporos Int* 2006;17:337–47.
95. Ward LM, Gaboury I, Ladhani M, Zlotkin S. Vitamin D-deficiency rickets among children in Canada. *Can Med Assoc J* 2007;177:161–6.
96. Stene LC, Ulriksen J, Magnus P, Joner G. Use of cod liver oil during pregnancy associated with lower risk of type I diabetes in the offspring. *Diabetologia* 2000;43:1093–8.
97. Chiu KC, Chu A, Go VL, Saad MF. Hypovitaminosis D is associated with insulin resistance and beta cell dysfunction. *Am J Clin Nutr* 2004;79:820–5.
98. Hypponen E, Laara E, Reunanen A, Jarvelin MR, Virtanen SM. Intake of vitamin D and risk of type 1 diabetes: a birth-cohort study. *Lancet* 2001;358:1500–3.
99. Sowers M. Pregnancy and lactation as risk factors for subsequent bone loss and osteoporosis. *J Bone Miner Res* 1996;11:1052–60.
100. Specker BL, Tsang RC, Ho ML. Changes in calcium homeostasis over the first year postpartum: effect of lactation and weaning. *Obstet Gynecol* 1991;78:56–62.

